

JC07 Rec'd PCT/PTO 19 FEB 2002

Form PTO-1390U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE (REV 10-95) TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US) CONCERNING A FILING UNDER 35 U.S.C. 371		ATTORNEY'S DOCKET NUMBER 3274-020104
		U.S. APPLICATION NO (If known, see 37 CFR 1.5) 10/049964
INTERNATIONAL APPLICATION NO PCT/JP00/05576	INTERNATIONAL FILING DATE 18.08.00 (18 August 2000)	PRIORITY DATES CLAIMED 19.08.99 (19 August 1999)
TITLE OF INVENTION METHOD FOR PRODUCING GLYCOSIDES		
APPLICANT(S) FOR DO/EO/US Nozomu YASUTAKE, Shinsuke MIYOSHI, Taiichi USUI, Takeomi MURATA and Kazuhide TOTANI		
<p>Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items And other information:</p> <ol style="list-style-type: none"> <input checked="" type="checkbox"/> This is a FIRST submission of items concerning a filing under 35 U.S.C. 371. <input type="checkbox"/> This is a SECOND or SUBSEQUENT submission of items concerning a filing under 35 U.S.C. 371. <input checked="" type="checkbox"/> This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1). <input checked="" type="checkbox"/> A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date <input checked="" type="checkbox"/> A copy of the International Application as filed (35 U.S.C. 371(c)(2)) <ol style="list-style-type: none"> <input type="checkbox"/> is transmitted herewith (required only if not transmitted by the International Bureau) <input checked="" type="checkbox"/> has been transmitted by the International Bureau. <input type="checkbox"/> is not required, as the application was filed in the United States Receiving Office (RO/US) <input checked="" type="checkbox"/> A translation of the International Application into English (35 U.S.C. 371(c)(2)) <input checked="" type="checkbox"/> Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3)) <ol style="list-style-type: none"> <input type="checkbox"/> are transmitted herewith (required only if not transmitted by the International Bureau). <input type="checkbox"/> have been transmitted by the International Bureau. <input type="checkbox"/> have not been made; however, the time limit for making such amendments has NOT expired <input checked="" type="checkbox"/> have not been made and will not be made. <input type="checkbox"/> A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)). <input type="checkbox"/> An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)). <input type="checkbox"/> A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)) <p>Items 11. to 16. below concern document(s) or information included:</p> <ol style="list-style-type: none"> <input type="checkbox"/> An Information Disclosure Statement under 37 CFR 1.97 and 1.98 <input type="checkbox"/> An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included <input checked="" type="checkbox"/> A FIRST preliminary amendment <input type="checkbox"/> A SECOND or SUBSEQUENT preliminary amendment <input type="checkbox"/> A substitute specification. <input type="checkbox"/> A change of power of attorney and/or address letter <input checked="" type="checkbox"/> Other items or information: <ol style="list-style-type: none"> WO 01/14575-Front Page (2 pp) And International Search Report (1 p.) 		

U.S. APPLICATION NO. (If known, file 37 CFR 1.53)		INTERNATIONAL APPLICATION NO PCT/JP00/05576		ATTORNEY'S DOCKET NUMBER 3274-020104	
17. <input checked="" type="checkbox"/> The following fees are submitted: BASIC NATIONAL FEE (37 CFR 1.492(a)(1)-(5)): Search Report has been prepared by the EPO or JPO \$890.00 International preliminary examination fee paid to USPTO (37 CFR 1.482) \$710.00 No international preliminary examination fee paid to USPTO (37 CFR 1.482) but international search fee paid to USPTO (37 CFR 1.445(a)(2)) \$740.00 Neither international preliminary examination fee (37 CFR 1.482) nor International search fee (37 CFR 1.445(a)(2)) paid to USPTO \$1040.00 International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(2)-(4) \$100.00 ENTER APPROPRIATE BASIC FEE AMOUNT =				CALCULATIONS PTO USE ONLY	
				\$ 890.00	
Surcharge of \$130.00 for furnishing the oath or declaration later than <input type="checkbox"/> 20 <input checked="" type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(e)).				\$ 130.00	
CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE		
Total claims	17 - 20	0	X \$18.00	\$ 0.00	
Independent claims	2 - 3 =	0	X \$84.00	\$ 0.00	
MULTIPLE DEPENDENT CLAIM(S) (if applicable)			+ \$280.00	\$ 0.00	
TOTAL OF ABOVE CALCULATIONS =				\$ 1020.00	
Reduction of 1/2 for filing by small entity, if applicable				\$ 0.00	
SUBTOTAL =				\$ 1020.00	
Processing fee of \$130.00 for furnishing the English translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(f)) +				\$ 0.00	
TOTAL NATIONAL FEE =				\$ 1020.00	
Fee for recording the enclosed assignment (37 CFR 1.21(h)) The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property +				\$ 0.00	
TOTAL FEES ENCLOSED =				\$ 1020.00	
				Amount to be: Refunded	\$
				Charged	\$
a <input checked="" type="checkbox"/> A check in the amount of \$ <u>1020</u> to cover the above fees is enclosed.					
b <input type="checkbox"/> Please charge my Deposit Account No. _____ in the amount of \$ _____ to cover the above fees A duplicate copy of this sheet is enclosed					
c <input checked="" type="checkbox"/> The Assistant Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. <u>23-0650</u> A duplicate copy of this sheet is enclosed.					
NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.					
SEND ALL CORRESPONDENCE TO: Kent E. Baldauf 700 Koppers Building 436 Seventh Avenue Pittsburgh, Pennsylvania 15219-1818 Telephone: (412) 471-8815 Facsimile: (412) 471-4094					
SIGNATURE <u>Kent E. Baldauf</u> NAME <u>25,826</u> REGISTRATION NUMBER					

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PATENT APPLICATION/PCT
Attorney Docket No. 3274-020104

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of :

Nozomu YASUTAKE : METHOD FOR PRODUCING GLYCOSIDES
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Taiichi USUI :
Takeomi MURATA :
Kazuhide TOTANI :

International Application :
No. PCT/JP00/05576 :

International Filing Date :
18 August 2000 :

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19 August 1999 :

Serial No. Not Yet Assigned :

Filed Concurrently Herewith :

Pittsburgh, Pennsylvania
February 19, 2002

PRELIMINARY AMENDMENT

Commissioner for Patents
Washington, D.C. 20231

Sir:

Prior to initial examination, please amend the above-identified patent application as follows:

IN THE SPECIFICATION:

On page 1, please delete the section heading "DESCRIPTION".

Please amend claim 6 as follows:

6. The method of claim 1, wherein the enzyme having an activity of cleaving a β -1,4 glucosyl bond is exo-cellobiohydrolase, β -D-glucosidase, and/or cellulase.

Please add new claims 8-17 as follows:

8. The method of claim 2, wherein X is hydrogen.

9. The method of claim 3, wherein X is hydrogen.

10. The method of claim 4, wherein X is hydrogen.

11. The method of claim 2, wherein the enzyme having an activity of cleaving a β -1,4 glucosyl bond is exo-cellobiohydrolase, β -D-glucosidase, and/or cellulase.

12. The method of claim 3, wherein the enzyme having an activity of cleaving a β -1,4 glucosyl bond is exo-cellobiohydrolase, β -D-glucosidase, and/or cellulase.

13. The method of claim 4, wherein the enzyme having an activity of cleaving a β -1,4 glucosyl bond is exo-cellobiohydrolase, β -D-glucosidase, and/or cellulase.

14. The method of claim 5, wherein the enzyme having an activity of cleaving a β -1,4 glucosyl bond is exo-cellobiohydrolase, β -D-glucosidase, and/or cellulase.

15. The method of claim 8, wherein the enzyme having an activity of cleaving a β -1,4 glucosyl bond is exo-cellobiohydrolase, β -D-glucosidase, and/or cellulase.

16. The method of claim 9, wherein the enzyme having an activity of cleaving a β -1,4 glucosyl bond is exo-cellobiohydrolase, β -D-glucosidase, and/or cellulase.

17. The method of claim 10, wherein the enzyme having an activity of cleaving a β -1,4 glucosyl bond is exo-cellobiohydrolase, β -D-glucosidase, and/or cellulase.

IN THE ABSTRACT:

On page 29, please replace the section heading "ABSTRACT" with the following rewritten section heading:

Abstract of the Disclosure

Please replace the first partial paragraph on page 29 with the following rewritten paragraph:

A lactosyl glycoside or an N-acetyllactosaminy glycoside useful as foods, functional food materials, medicines, and reagents is provided with high yield and inexpensively by using a transfer reaction represented by the following formula in the presence of an enzyme having an activity of cleaving a β 1,4 glucosyl bond:

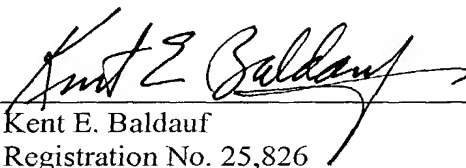
REMARKS

The specification and claims have been amended to place the application in conformance with standard United States patent practice.

Examination and allowance of pending claims 1-17 are respectfully requested.

Respectfully submitted,

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VERSION WITH MARKINGS TO SHOW CHANGES MADE

In the specification:

On page 1, the section heading "Technical Field" has been amended as follows:

[Technical Field] Background of the Invention

1. Field of the Invention

On page 1, the section heading "Background Art" has been amended as follows:

[Background Art] 2. Description of the Prior Art

On page 5, the section heading "Disclosure of Invention" has been amended as follows:

[Disclosure of Invention] Summary of the Invention

On page 6, the third complete paragraph has been amended as follows:

Figure 2 is a ¹H-NMR analysis of ethyl lactoside.

On page 6, the fourth complete paragraph has been amended as follows:

Figure 3 is a mass spectrum analysis of ethyl lactoside.

On page 7, the section heading "Best Mode for Carrying Out the Invention" has been amended as follows:

[Best Mode for Carrying Out the Invention] Detailed Description of the Invention

In the claims:

Claim 5 has been amended as follows:

5. The method of [any one of claims 1 to 4] claim 1, wherein X is hydrogen.

Claim 6 has been amended as follows:

6. The method of [any one of claims 1 to 5] claim 1, wherein the enzyme having an activity of cleaving a β -1,4 glucosyl bond is exo-cellobiohydrolase, β -D-glucosidase, and/or cellulase.

In the abstract:

On page 29, the section heading "ABSTRACT" has been amended as follows:

[ABSTRACT] Abstract of the Disclosure

On page 29, the first partial paragraph has been amended as follows:

[The present invention provides a] A lactosyl glycoside or an N-acetyllactosaminyl glycoside useful as foods, functional food materials, medicines, and reagents is provided with high yield and inexpensively by using a transfer reaction represented by the following formula in the presence of an enzyme having an activity of cleaving a β 1,4 glucosyl bond:

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2001 年 3 月 1 日 (01.03.2001)

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(81) 指定国 (国内): AU, CA, JP, KR, US.

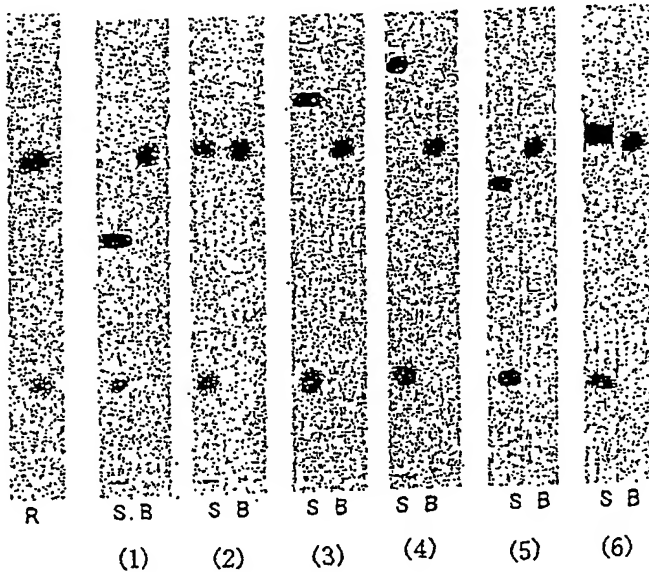
[続葉有]

(54) Title: PROCESS FOR PRODUCING GLYCOSIDES

(54) 発明の名称: 配糖体の製造方法

Lac-pNP→

Lac→



(57) Abstract: A process for efficiently and economically producing a lactosyl glycoside and an N-acetyllactosaminyl glycoside, which are useful as foods, functional food materials, drugs and reagents, by using a transfer reaction represented by the following general formula in the presence of an enzyme having an activity of cleaving a β 1,4 glycosyl bond: $\text{LacA-X} + \text{Y} \rightarrow \text{LacA-Y} + \text{X}$ wherein LacA represents lactose or N-acetyllactosamine; X represents hydrogen (H), a saccharide, a glycoconjugate or a phenolic compound; and Y represents a compound having alcoholic hydroxy or phenolic hydroxy.

[続葉有]



WO 01/14575 A1

5/p, 1/2

DESCRIPTION

Method for producing glycosides

Technical Field

The present invention relates to a method for producing lactosyl glycosides or N-acetyllactosaminyl glycosides. More specifically, the present invention relates to a method for producing lactosyl glycosides or N-acetyllactosaminyl glycosides, using an enzyme having an activity of cleaving a β -1,4 glucosyl bond.

Background Art

Oligosaccharides and glycoconjugates (glycoproteins, glycolipids, glycosides, etc.) are present in animals and plants, for example, biological cells, body fluids, fruits, seeds, etc. These components have essential functions to maintain or regulate biological functions such as the transmission of biological information on the surface of a cell membrane, the morphogenesis of cells, the maintenance of a specific structure that is composed of covalent bonds with proteins, just like genes and proteins.

Glycolipids, certain kinds of glycoconjugates, are classified into sphingoglycolipids, glykoglycerolipids and others. Sphingoglycolipids are present mostly in animal tissues, and are classified roughly into four series, namely the globo-, the lacto-, the ganglio-, and the gala-series, depending on their basic sugar chain component. Three of those series, the globo-, the lacto-, and the ganglio-series, have a lactose unit and lactosylceramide unit in which lactose is bound to ceramide as the basic backbone, and are classified depending on the saccharide to be added next.

Lactosylceramide has a Gal β 1-4Glc unit, that is, a lactose unit. It has come to be known that bacteria recognize this lactose unit to recognize the sequence of a glycoconjugate. The mechanism that a microorganism adheres to a host cell via a sugar chain plays an important role as a model of inter-cell recognition. Furthermore, it has come to be known that bacteria not only recognize the end of a sugar chain on the surface of a cell for binding, but also recognize the sugar chain structure inside the sugar chain. For example, it is known that *E. coli*, microorganisms belonging to genus *Pseudomonas*, *Actinomycetes*, and propionibacteria recognize the lactose unit that is present inside of various sugar chains as well.

Thus, lactosylceramide as described above plays a physiologically important role in such as adhesion of cells. Therefore, lactosyl glycoside can be used for the applications of medicine, food and the like, but it is not easy to produce the lactosyl glycoside at present.

There are many kinds of glycosides as glycoconjugates. For example, the glycosides which are composed of sugar unit (glucose, galactose and N-acetyllactosaminide), and glycerol as the aglycon, are known. Naturally occurring components such as glyceroyl glucoside (2-O- α -D-Glucosylglycerol, (2R)-1-O- α -D-Glucosylglycerol, and (2S)-1-O- α -D-Glucosylglycerol) are known as the glycoside for glucose. Glyceroyl galactoside (1-O- β -D-Galactosylglycerol) as a chemically synthesized substance is known as the glycoside for galactose. As the glycoside for N-acetyllactosaminide, the existence of O- β -D-Galactopyranosyl-(1 \rightarrow 4)-O- β -D-2-Acetyl-amino-2-deoxyl-Glucopyranosyl-(1 \rightarrow 2)-Glycerol is reported.

Among the above-described glycosides, glyceroyl glucoside, glyceroyl galactoside and the like are characterized by their taste, their moisture retaining properties, and their physiological functions.

These substances are also useful as a starting material for the synthesis of glyco glycerolipids. Glyco glycerolipids are present widely in organisms, ranging from microorganisms to higher plants, and are reported to have emulsification stability and carcinogenic promoter suppressing effects. Therefore, compounds in which saccharides having different structures are added to glycerol can be expected to have new functions or to be applied as new starting materials of glyco glycerolipids.

N-Acetyl lactosaminyl glycosides are known to be present in a receptor recognized by Staphylococcus or the core unit of the sugar chain structure recognized by influenza viruses A and B, Sendai virus, Newcastle disease virus and the like, and participate in the recognition of cells. Therefore, the N-acetyl lactosaminyl glycosides are useful for medical applications, but it is not easy to produce N-acetyl lactosaminyl glycosides at present.

The functions of such saccharide-related substances are noted, therefore, modification and substitution research (remodeling) of sugar chains and glyco conjugates is performed in order to utilize the substances for medical and food applications by improving these functions further or modifying their physiological functions.

For remodeling the sugar chains of saccharides and glyco conjugates, there are chemical methods, enzymatic methods, and methods combining chemical and enzymatic methods.

As a chemical method, there is an organic synthesis method for sugar chain synthesis, but this method involves many complicated steps, and a process for removing byproducts (purifying a desired substance) is necessary. Furthermore, reagents that are harmful to human health are often used, and there is the fear that the waste liquid may damage the surrounding environment.

On the other hand, for remodeling sugar chains enzymatically, a method using transferase, exoglucosidase, endoglucosidase, and the like are known.

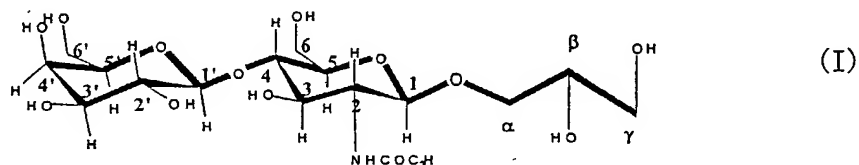
One of the methods using transferase or exoglucosidase is described in D. H. Joziase et al. *Eur. J. Biochem.*, vol. 191, pages 75-83 (1990), which discloses a method of sequentially adding a glucosyl group to a nonreducing end of a sugar chain by using exoglucosidase or glucosyltransferase.

However, although the sugar chain synthesis using exoglucosidase or glucosyltransferase is easier than chemical synthesis, it is still necessary to perform an enzyme reaction sequentially for each saccharide residue one by one, so that a large number of reaction steps are required, which makes the method troublesome.

Examples of the disclosure of transglycosylation using endoglucosidase are as follows. R. B. Trimble et al. *J. Biol. Chem.*, vol. 261, pp. 12000-12005 (1986) describe a method of using endo- β -N-acetylglucosaminidase derived from *Flavobacterium meningosepticum*. R. M. Bardales et al., *J. Biol. Chem.*, vol. 264, pp. 19893-19897 (1989) describe a method of using endo- α -N-acetylgalactosaminidase derived from *Diprococcus pneumoniae*. Japanese Laid-Open Patent Publication (Tokkai) No. 5-64594 describes a transfer reaction of high-mannose type of sugar chains to saccharides by using endo- β -N-acetylglucosaminidase derived from *Arthrobacter protophormiae*. K. Yamamoto et al., *Biochem. Biophys. Res. Commun.*, vol. 203, pp. 244-252 (1994) describe a transfer reaction of a sugar chain to a saccharide by using endo- β -acetylglucosaminidase derived from *Mucor hiemalis*. Japanese Laid-Open Patent Publication (Tokkai) No. 10-245402 describes a transfer reaction of (Man)₆-GlcNAc using endo- β -N-acetylglucosaminidase. Japanese Laid-Open Patent Publication

cleaving a β -1,4 glucosyl bond is exo-cellobiohydrolase, β -D-glucosidase, and/or cellulase.

The present invention is also directed to glyceroyl N-acetyllactosaminide (O- β -D-Galactopyranosyl-(1 \rightarrow 4)-O- β -D-2-Acetyl-amino-2-deoxyl-Glucopyranosyl-(1 \rightarrow 1)-Glycerol) represented by the structural formula (I):



Brief Description of Drawings

Figure 1 is the results of thin layer chromatography showing a transfer reaction of the lactosyl group of Lac-pNP to an aliphatic alcohol.

Figure 2 is ^1H -NMR of ethyl lactoside.

Figure 3 is a mass spectrum of ethyl lactoside.

Figure 4 is the results of thin layer chromatography showing a transfer reaction of the lactosyl group of Lac-pNP to an amino acid.

Figure 5 is the results of thin layer chromatography showing a transfer reaction of the lactosyl group of Lac-pNP to a monosaccharide.

Figure 6 is the results of thin layer chromatography showing a transfer reaction of the lactosyl group of Lac-pNP to L-menthol.

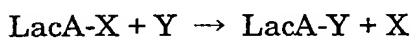
Figure 7 is the results of thin layer chromatography showing a transfer reaction of the N-acetyllactosaminyl group of LacNAc-pNP to an aliphatic alcohol.

Figure 8 is the results of thin layer chromatography showing a condensation reaction of lactose and an aliphatic alcohol or a monosaccharide.

Figure 9 is the results of thin layer chromatography showing a condensation reaction of N-acetyllactosaminide and an aliphatic alcohol or a monosaccharide.

Best Mode for Carrying Out the Invention

The present invention is directed to a method for producing a lactosyl glycoside or an N-acetyllactosaminyl glycoside characterized by using a transfer reaction represented by the following formula in the presence of an enzyme having an activity of cleaving a β 1,4 glucosyl bond:



(Enzyme)

There is no limitation regarding the enzymes used in the present invention, as long as they have an activity of cleaving a β 1,4 glucosyl bond. Either endo-type or exo-type enzyme can be used. Preferable examples of the enzymes include exo-cellobiohydrolase, β -D-glucosidase, and cellulase. These enzymes can be used alone or in combination.

Commercially available enzymes having an activity of cleaving a β 1,4 glucosyl bond can be also used in the present invention. Microorganisms that produce the enzymes having an activity of cleaving a β 1,4 glucosyl bond, culture medium or extracts thereof, or purified products can be used in the present invention. Microorganisms that produce the enzymes having an activity of cleaving a β 1,4 glucosyl bond are known.

Examples of bacteria include microorganisms belonging to genus Bacillus, Cellvibrio, Cellulomonas, Pseudomonas, Sporocytophaga, Acetivibrio, Clostridium, Bacterioides, Treponema,

and Ruminococcus.

Examples of filamentous fungi include fungi belonging to genus Humicola, Trichoderma, Myrothecium, Aspergillus, Irpex, Penicillium, and Pellicularia.

Examples of actinomyces include microorganisms belonging to genus Streptomyces, and Thermonospola.

(Donor)

Examples of the lactose or N-acetyllactosaminide donor represented by the formula LacA-X include lactose or N-acetyllactosaminide (when X is hydrogen), and saccharides, glycoconjugates or phenolic compounds containing lactose or N-acetyllactosaminide.

Examples of the saccharides as the lactose donor and/or the N-acetyllactosaminide donor include oligosaccharides containing lactose or N-acetyllactosaminide connected with a β -glucoside linkage.

Examples of the glycoconjugates as the lactose donor and/or the N-acetyllactosaminide donor include glycoproteins, proteoglycans, and glycolipids having lactose or N-acetyllactosaminide at their end. As the glycolipids, sphingoglycolipids are preferable, and one having lactose or N-acetyllactosaminide at the end of a ceramide, which is a component of sphingoglycolipid, or an analogue of the ceramide is preferably used.

There is no particular limitation regarding the phenolic compounds as the lactose or N-acetyllactosaminide donor. Examples thereof include phenol and its derivatives (e.g., para-nitrophenol), tyrosine and its derivatives, salicylic acid and its derivatives having lactose or N-acetyllactosaminide. When LacA-X is lactose or N-acetyllactosaminide, a condensation reaction occurs between LacA-X

and an acceptor Y, and this reaction also is included in the transfer reaction referred to in the present invention.

(Acceptor)

Compounds having a hydroxyl group, that is, an alcoholic hydroxyl group or a phenolic hydroxyl group can be used as Y, which is the acceptor of lactose or N-acetyllactosaminide.

Examples of the compounds having an alcoholic hydroxyl group as the acceptor of lactose or N-acetyllactosaminide include aliphatic alcohols, amino acids, and saccharides.

Aliphatic alcohols have a structure of aliphatic hydrocarbon in which one or more hydrogen atom(s) is substituted with a hydroxyl group, and can be classified into primary alcohols, secondary alcohols, and tertiary alcohols, depending on the number of alkyl groups binding to a carbon atom bound to the hydroxyl group. They can be classified into monohydric alcohols, dihydric alcohols and polyhydric alcohols, depending on the number of binding of hydroxyl group.

Examples of primary alcohols include methanol, ethanol, butanol, propanol, hexanol (cis-3-hexanol), octanol, allyl alcohol, geraniol, and citronellol.

Examples of secondary alcohols include isopropanol, and sec-butanol.

Examples of tertiary alcohols include t-butylalcohol.

A typical example of a monohydric alcohol is methanol, a typical example of a dihydric alcohol is ethylene glycol, and a typical example of a trihydric alcohol is glycerol.

In addition to the above-described substances, alicyclic alcohols having a ring structure such as menthol and santalol are included in the aliphatic alcohols.

Examples of the amino acids having an alcoholic hydroxyl group include serine and threonine. Furthermore, peptides and proteins containing the residue of an amino acid having an alcoholic hydroxyl group such as serine and threonine, or peptides containing tyrosine having a phenolic hydroxyl group as described later can be also used as the acceptor of the present invention.

Examples of the saccharides having an alcoholic hydroxyl group include monosaccharides such as glucose and mannose. Examples of the oligosaccharides include: starch-related oligosaccharides such as maltooligosaccharides, isomaltooligosaccharides, cyclodextrin, gentiooligosaccharides and nigerooligosaccharides; sugar-related oligosaccharides such as maltooligosylsucroses, fructooligosaccharides, paratinose, lactosucrose, xylosylfructoside, raffinose and stachyose; lactose-related oligosaccharides such as galactooligosaccharides, lactosucrose and lactulose; xylooligosaccharides; agarooligosaccharides; chitin/chitosan oligosaccharides; mannoooligosaccharides; and alginate oligosaccharides. Furthermore, a sugar chain that is a component of a glycoprotein or a glycolipid, or a sugar chain that is a partial structure thereof can be used.

Examples of the compounds having a phenolic hydroxyl group include phenol and its derivatives, tyrosine and its derivatives, salicylic acids and their derivatives, and soybean isoflavones such as genistein.

In addition, glycoconjugates such as glycoproteins and sphingoglycolipids, conjugated lipids such as ceramide, flavonoid analogues such as flavonol have a hydroxyl group, and can be used as the acceptor.

(Lactosyl or N-acetyllactosaminyl transfer reaction)

There is no particular limitation regarding the lactosyl or N-

acetyllactosaminyl transfer reaction. It is sufficient to mix a lactosyl or N-acetyllactosaminyl donor, an enzyme having an activity of cleaving a β -1,4 glucosyl bond, and a lactosyl or N-acetyllactosaminyl acceptor in an appropriate ratio to allow to react.

There is no particular limitation regarding the concentrations of the lactosyl or N-acetyllactosaminyl donor and acceptor in the transfer reaction, and the concentrations can be determined as appropriate by those skilled in the art, in view of the efficiency of the reaction or the like.

There is no particular limitation regarding the concentration of the enzyme used, and the concentration can be determined as appropriate by those skilled in the art, in view of the concentration of the substrate, the reaction temperature, the reaction time or the like. For example, the amount of the enzyme to be used can be determined by performing a preliminary test.

There is no particular limitation regarding the reaction temperature. In general, the reaction temperature is in the range of about $\pm 20^{\circ}\text{C}$ of the optimum temperature of the enzyme used. More preferably, it is in the range of about $\pm 10^{\circ}\text{C}$ of the optimum temperature of the enzyme.

There is no particular limitation regarding the pH for the reaction. Although it depends on the enzyme used, in general, the reaction is performed in the range of about ± 3 of the optimum pH of the enzyme used, more preferably about ± 2 of the optimum pH of the enzyme. For pH adjustment to such a value, a suitable buffer routinely used by those skilled in the art can be used.

When it is difficult to dissolve the substrate in water, a small amount of a solvent or a surfactant can be added to water to dissolve the substrate to increase the reaction efficiency.

Examples

Hereinafter, the present invention will be described by way of examples, but the present invention is not limited thereto.

Example 1

A lactosyl transfer reaction was performed by using aliphatic alcohols as the acceptor. As the lactose donor, p-nitrophenyl- β -D-lactoside (hereinafter, referred to as "Lac-pNP") was used, and ethanol, butanol, hexanol, octanol, isopropanol, and t-butylalcohol, which are aliphatic alcohols, were used as the lactose acceptor. As the enzyme, cellulase (manufactured by Kyowa Hakko Kogyo Co., Ltd.) derived from *Trichoderma reesei* was used.

First, 200 μ l of 4.65 mg/ml of Lac-pNP, 120 μ l of one of the alcohols, and 30 μ l of 0.2M acetate buffer (pH 5.5) were added to 50 μ l of 10 mg/ml of an enzyme solution (dissolved in 20 mM acetate buffer (pH 5.0)) so that the total volume was 400 μ l. An enzyme reaction was performed at 40°C, and a transfer reaction was monitored over time by thin layer chromatography (hereinafter, referred to as "TLC": with chloroform: methanol: water = 60: 35: 8 as the developing solvent). Aluminum Sheet Silicagel 60/Kieselgur F254 Precoat manufactured by Merck was used for the TLC. Detection was performed by color-development with orcin-sulfuric acid method.

The results are shown in Figure 1. In Figure 1, R shows a mixture of lactose (Lac) and Lac-pNP, and the R_f value (0.13) of the lactose is smaller than that (0.43) of Lac-pNP. S refers to a reaction mixture, and B refers to a blank. The blank was obtained by adding water instead of the enzyme for reaction. In Figure 1, (1), (2), (3), (4), (5) and (6) show the results when ethanol, butanol, hexanol, octanol, isopropanol, and t-butylalcohol, respectively, were used as the

substrate.

Lactose (Lac) that had not been present in an early stage of the reaction was produced in all the reaction mixtures S of (1) to (6), and alkyl lactoside corresponding to each substrate of (1) to (6) (ethyl lactoside, butyl lactoside, hexyl lactoside, octyl lactoside, isopropyl lactoside, and t-butyl lactoside, respectively) was further produced. In other words, it was confirmed that in all of the reaction mixtures of (1) to (6), the lactosyl group of Lac-pNP was transferred to the hydroxyl group of the aliphatic alcohol, so that the alkyl lactoside was produced. The R_f value (0.38) of the butyl lactoside of (2) and the R_f value (0.45) of the t-butyl lactoside of (6) were close to the R_f value (0.43) of Lac-pNP, so that in the TLC of Figure 1, (2) and (6) were difficult to distinguish from Lac-pNP visually.

Example 2

The reaction product ((1)) obtained using ethanol as the lactose acceptor of Example 1 was analyzed. A reaction was performed for 27 hours, and the reaction product was heated at 100°C for 10 minutes, and centrifuged at 17,000 rpm for 10 minutes to give a supernatant. The supernatant was evaporated to dryness, and then dissolved in 10 ml of 25% aqueous methanol. The solution was loaded to Toyopearl HW-40S gel filtration column (ϕ 2.5 × 80 cm), and fractionated into 10ml fractions with a 25% aqueous methanol as the eluent at a flow rate of 1.0 ml per min. The fractions were detected by measuring the UV absorption at 210 nm and 300 nm and the color-development (485 nm) by the phenol-sulfuric acid method. The fractions of tube Nos. 34 to 44 were pooled and evaporated to dryness.

The dried product was dissolved in 10 ml of chloroform: methanol: water = 50: 45: 10 to separate and purify by silica gel column

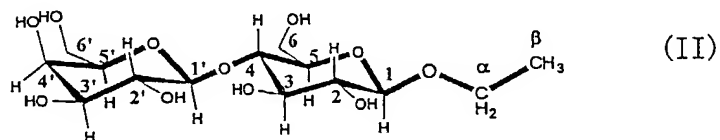
chromatography using Wakogel C-300. Elution was performed at a flow rate of 2.0 ml/min with chloroform: methanol: water = 50: 45: 10 as the developing solvent, and the eluate was fractionated into 10 ml fractions. The fractions were detected by measuring the UV absorption at 210 nm and 300 nm and the color-development (485 nm) by the phenol-sulfuric acid method. The fractions of tube Nos. 11 to 13 were pooled and evaporated to dryness.

A portion of the obtained dried product was dissolved in 0.3 ml of distilled water and subjected to mass analysis FAB/MAS. On the other hand, a portion of the obtained dried product was dissolved in heavy water and subjected to ^1H -NMR analysis. The result of the ^1H -NMR is shown in Figure 2, and the result of the FAB/MAS is shown in Figure 3. The data of the ^1H -NMR and the FAB/MAS corresponding to Figures 2 and 3 are shown below.

^1H -NMR (270MHz, D_2O): δ 4.50(d, J 7.9Hz, H-1), 4.45(d, J 7.7Hz, H-1'), 3.30(dd, H-2), 1.24(t, H- β);

FAB/MAS: (m/z) = 371 ($\text{C}_{14}\text{H}_{26}\text{NO}_{11}+\text{H}$) $^+$, (m/z) = 393 ($\text{C}_{14}\text{H}_{26}\text{NO}_{11}+\text{Na}$) $^+$.

These data confirmed that the product of (1) was ethyl lactoside represented by the structural formula (II):



Example 3

A lactosyl transfer reaction to an amino acid having an alcoholic hydroxyl group was performed. The same enzyme and lactose donor as that in Example 1 were used. As a reaction mixture, 400 μl of 15 mM acetate buffer (pH 5.0) containing 0.5 mg of the enzyme, 0.94 mg of

Lac-pNP, and 50 mg of serine or 15 mg of threonine were prepared, and allowed to react at 40°C for 24 hours. After the reaction, analysis was performed by TLC in the same manner as in Example 1. The results are shown in Figure 4. The developing solvent was chloroform: methanol: water = 50: 45: 10.

In Figure 4, R shows a mixture of Lac and Lac-pNP, S shows a reaction mixture, and B shows a blank. The results when serine was used as the acceptor is shown in (7), and the results when threonine was used as the acceptor is shown in (8). In the reaction mixtures of (7) and (8), the substrate Lac-pNP disappeared and seryl lactoside and threonyl lactoside were produced, respectively. The R_f values of the seryl lactoside and the threonyl lactoside were 0.27 and 0.29, respectively, and were close to the R_f value (0.34) of lactose so that the seryl lactoside and the threonyl lactoside were not distinctly separated from lactose but they appeared as a broad band.

Example 4

A lactosyl transfer reaction to a monosaccharide having an alcoholic hydroxyl group was performed. The same enzyme and lactose donor as in Example 1 were used. As a reaction mixture, 400 µl of 15 mM acetate buffer (pH 5.0) containing 0.5 mg of the enzyme, 0.94 mg of Lac-pNP, and 70 mg of glucose or mannose were prepared, and allowed to react at 40°C for 24 hours. After the reaction, analysis was performed by TLC in the same manner as in Example 1. The results are shown in Figure 5. The developing solvent was chloroform: methanol: water = 50: 45: 10.

In Figure 5, R shows a mixture of Lac and Lac-pNP, S shows a reaction mixture, and B shows a blank. The results when glucose was used as the acceptor is shown in (9), and the results when mannose was

used as the acceptor is shown in (10). In the reaction mixtures of (9) and (10), the substrate Lac-pNP disappeared and glucosyl lactoside and mannosyl lactoside were produced, respectively. The R_f values of the glucosyl lactoside and the mannosyl lactoside were 0.24 and 0.25, respectively, and were smaller than the R_f value (0.34) of lactose (Lac) so that they were distinctly separated from lactose.

In Figure 5, there are bands between Lac-pNP and Lac. In (9), the band is glucose (Glu) and in (10), the band is mannose (Man).

Example 5

A lactosyl transfer reaction to L-menthol, which is one of alicyclic alcohols having an alcoholic hydroxyl group, was performed. The same enzyme and lactose donor as that in Example 1 were used. As a reaction mixture, 400 μ l of 15 mM acetate buffer (pH 5.0) containing 1 mg of the enzyme, 1 mg of Lac-pNP, 5 mg of L-menthol, and 2 wt% of cholic acid as a surfactant were prepared, and allowed to react at 40°C for 24 hours. After the reaction, analysis was performed by TLC in the same manner as in Example 1. The results are shown in Figure 6. The developing solvent was chloroform: methanol: water = 65: 35: 4.

In Figure 6, R shows a mixture of Lac and Lac-pNP, S shows a reaction mixture, and B shows a blank. The substrate Lac-pNP disappeared, and L-mentholyl lactoside (Lac-menthol) was produced (R_f value of 0.62) and was distinguished from lactose (R_f value of 0.13). The spots other than that of L-mentholyl lactoside and the cholic acid having an R_f value of 0.81 seem to indicate some reaction products.

Example 6

An N-acetyllactosaminyl transfer reaction to an aliphatic

alcohol having an alcoholic hydroxyl group was performed.

As the N-acetyllactosaminide donor, p-nitrophenyl- β -N-acetyllactosaminide (hereinafter, referred to as "LacNAc-pNP") was used. As a reaction mixture, 400 μ l of 15 mM acetate buffer (pH 5.0) containing 0.94mg of LacNAc-pNP, 120 μ l of methanol, ethanol or propanol as the N-acetyllactosaminide (hereinafter, referred to as "LacNAc") acceptor, 0.5 mg of cellulase (manufactured by Kyowa Hakko Co., Ltd.) derived from *Trichoderma reesei* as the enzyme were prepared, and allowed to react at 40°C for 24 hours. After the reaction, analysis was performed by TLC in the same manner as in Example 1. The results are shown in Figure 7. The developing solvent was chloroform: methanol: water = 50: 40: 50.

In Figure 7, R shows LacNAc and LacNAc-pNP, and B shows a blank. The substrate LacNAc-pNP (R_f =0.56) disappeared, and LacNAc (R_f value of 0.30) and new spots indicated by the arrows having R_f values of 0.38, 0.46, and 0.56 appeared, and they were found to correspond to methyl N-acetyllactosaminide (1), ethyl N-acetyllactosaminide (2) and propyl N-acetyllactosaminide (3), respectively. Among them, the spot of the ethyl N-acetyllactosaminide (2) by TLC matched the spot of a specimen (manufactured by Carbiochem).

Example 7

A lactosyl transfer reaction (condensation of lactose) using an aliphatic alcohol and a saccharide having an alcoholic hydroxyl group as an acceptor was performed. As the donor, lactose was used. As the acceptor, methanol, ethanol, propanol, butanol, hexanol, octanol, dodecanol, allyl alcohol, and mannose were used.

First, 650 μ l of 10 mM acetate buffer (pH 5.0) containing 100

$^1\text{H-NMR}$ (270 MHz, D_2O): δ 4.50(d, J 7.9Hz, H-1), 4.45(d, J 7.7Hz, H-1'), 3.30(dd, H-2), 1.24(t, H- β);

$^{13}\text{C-NMR}$ (270 MHz, D_2O): 17.1 (C- β), 62.9 (C-6), 63.8 (C-6'), 69.0(C-4'), 71.4 (C- α), 73.8 (C-2'), 75.4 (C-3'), 75.6 (C-2), 77.3 (C-3), 77.6 (C-5), 78.2 (C-5'), 81.3 (C-4), 104.5 (C-1), 105.7 (C-1').

From the above, the product (2) was confirmed to be ethyl lactoside represented by the structural formula (II).

7-2 Butyl lactoside (4), hexyl lactoside (5), octyl lactoside (6)

The spots of butyl lactoside (4), hexyl lactoside (5), and octyl lactoside (6) by TLC matched those of the transfer reaction product of Lac-pNP and the corresponding aliphatic alcohols obtained in Example 1.

Furthermore, the reaction mixture that seemed to contain octyl lactoside (6) was applied to the silica gel column (ϕ 2 \times 50 cm) as described above and eluted at a flow rate of 2.0 ml/min with chloroform : methanol : water = 60 : 25 : 4 as an eluent. The eluate was fractionated into 10 ml fractions and the reaction product was detected by color-development (485 nm) by the phenol-sulfuric acid method. Fractions Nos. 21 to 28 were collected and lyophilized to give about 5 mg of the reaction product. The purity of this product was confirmed to be 99% or more by TLC and liquid chromatography.

A portion of the obtained dried product was analyzed by FAB/MAS and the other portion of the obtained dried product was analyzed by $^1\text{H-NMR}$ and $^{13}\text{C-NMR}$. The results are shown below.

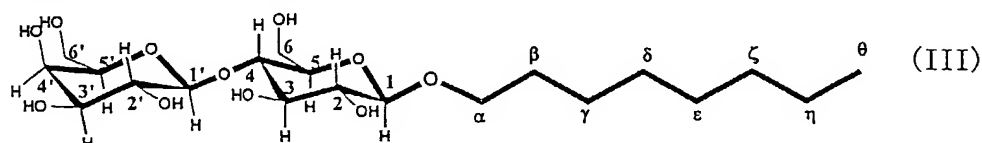
FAB/MAS: (m/z) = 455 ($\text{C}_{20}\text{H}_{38}\text{NO}_{11}+\text{H}$) $^+$, (m/z) = 477 ($\text{C}_{20}\text{H}_{38}\text{NO}_{11}+\text{Na}$) $^+$;

$^1\text{H-NMR}$ (270 MHz, D_2O): δ 4.48(d, J 8.0Hz, H-1), 4.45(d, J 7.7Hz, H-1'), 3.31(dd, H-2), 1.63(m, H- β), 1.36(m, H- γ), 1.29(m, H- δ -

η), 0.87(t, H- θ);

^{13}C -NMR (270 MHz, D_2O): 16.2 (C- θ), 24.8 (C- η), 27.8 (C- γ), 31.2 (C- δ), 31.2 (C- ϵ), 31.5 (C- β), 33.9 (C- ζ), 63.0 (C-6), 63.8 (C-6'), 71.4 (C-4'), 73.6 (C- α), 73.8 (C-2'), 75.4 (C-3'), 75.7 (C-2), 77.3 (C-3), 77.6 (C-5), 78.2 (C-5'), 81.3 (C-4), 104.8 (C-1), 105.7 (C-1').

From the above, the product (6) was confirmed to be octyl lactoside represented by the structural formula (III):



7-3 Dodecyl lactoside (7)

The reaction mixture that seemed to contain dodecyl lactoside (7) was applied to ODS column (ϕ 3.5 \times 60 cm) using Chromatorex-ODS DM1020T. The column was washed with 65% methanol in five volume of the column, and then eluted with 75% methanol (a flow rate of 2.5 ml/min). Fractionation was performed at 30 ml/fraction, and the reaction product was detected by color-development (485 nm) by the phenol-sulfuric acid method. Fractions Nos. 17 to 23 were collected and lyophilized to give about 5 mg of the reaction product. The purity of this product was confirmed to be 99% or more by TLC and liquid chromatography.

A portion of the obtained dried product was analyzed by FAB/MAS and the other portion of the obtained dried product was analyzed by ^1H -NMR and ^{13}C -NMR. The results are shown below.

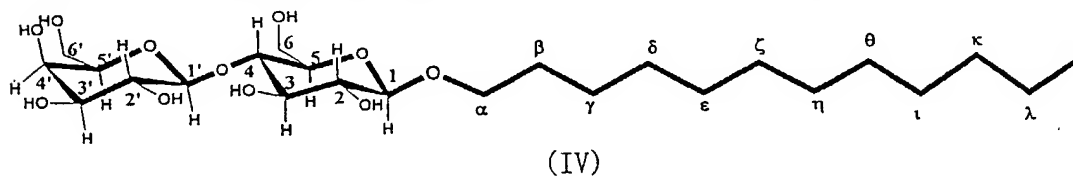
FAB/MAS: (m/z) = 511 ($\text{C}_{24}\text{H}_{46}\text{NO}_{11}+\text{H}$) $^+$, (m/z) = 533 ($\text{C}_{24}\text{H}_{46}\text{NO}_{11}+\text{Na}$) $^+$;

^1H -NMR (270 MHz, D_2O): δ 4.23(d, J 8.0Hz, H-1), 4.45(d, J 7.3Hz, H-1'), 3.19(dd, H-2), 1.57(m, H- β), 1.33(m, H- γ), 1.24(m, H- δ

$\sim \lambda$), 0.85(t, H- μ);

^{13}C -NMR (270 MHz, D_2O): 14.4 (C- μ), 23.7 (C- λ), 27.1 (C- γ), 30.5 (C- δ), 30.5 (C- ϵ), 30.6 (C- β), 30.8 (C- ζ), 30.8 (C- η), 30.8 (C- θ), 30.8 (C- τ), 33.1 (C- κ), 61.9 (C-6), 62.5 (C-6'), 70.3 (C-4'), 71.0 (C- α), 72.6 (C-2'), 74.8 (C-3'), 74.8 (C-2), 76.4 (C-3), 76.5 (C-5), 77.1 (C-5'), 80.7 (C-4), 104.3 (C-1), 105.1 (C-1').

From the above, the product (7) was confirmed to be dodecyl lactoside (7) represented by the structural formula (IV):



7-4 Mannosyl lactoside (8)

The spot of mannosyl lactoside (8) by TLC matched the spot of the mannosyl lactoside of the transfer product obtained in Example 4.

Example 8

An N-acetyllactosaminyl transfer reaction (condensation of N-acetyllactosaminide) to an aliphatic alcohol and a saccharide having an alcoholic hydroxyl group was performed.

Using N-acetyllactosaminide (LacNAc) as the donor and methanol, ethanol, propanol, and mannose as the acceptor, a reaction was performed under the same conditions as in Example 7. The results are shown in Figure 9. In Figure 9, S shows the reaction mixture, and B shows a blank. In all the reaction mixtures, a plurality of spots were seen, and new spots indicated by arrows having R_f values of 0.36, 0.40, and 0.45 appeared with respect to LacNAc having a R_f value = 0.24 (the developing solvent was chloroform: methanol: water = 60: 35: 8). These spots suggested that methyl N-acetyllactosaminide

(1), ethyl N-acetyllactosaminide (2), and propyl N-acetyllactosaminide (3) were produced ((1) to (3) in Figure 9).

With respect to the Rf value of 0.58 of LacNAc, a new spot having Rf values of 0.3 (the developing solvent was chloroform: methanol: water = 50: 45: 10) were seen. This spot suggested that mannosyl N-acetyllactosaminide (4) was produced ((4) in Figure 9).

8-1 Characterization of ethyl N-acetyllactosaminide (2)

The reaction mixture that seemed to contain ethyl N-acetyllactosaminide (2) was applied to the silica gel column (ϕ 2 \times 40 cm) as described above and eluted at a flow rate of 2.0 ml/min with chloroform : methanol : water = 50 : 45 : 10 as an eluent. The eluate was fractionated into 5 ml fractions and the reaction product was detected by color-development (485 nm) by the phenol-sulfuric acid method. Fractions Nos. 11 to 14 were collected and lyophilized to give about 5 mg of the reaction product. The purity of this product was confirmed to be 99% or more by TLC and liquid chromatography.

A portion of the obtained dried product was analyzed by FAB/MAS and the other portion of the obtained dried product was analyzed by ^1H -NMR and ^{13}C -NMR. The results are shown below.

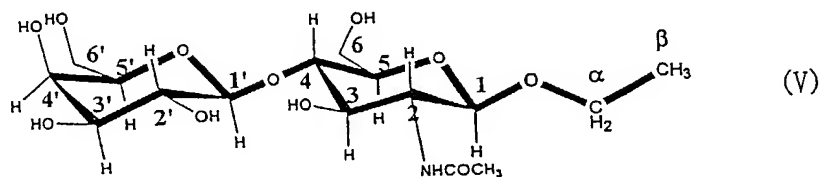
FAB/MAS: (m/z) = 411 ($\text{C}_{16}\text{H}_{29}\text{NO}_{11}+\text{H}$) $^+$, (m/z) = 433 ($\text{C}_{16}\text{H}_{29}\text{NO}_{11}+\text{Na}$) $^+$;

^1H -NMR (270 MHz, D_2O): δ 4.53(d, J 5.6Hz, H-1), 4.45(d, J 7.9Hz, H-1'), 2.02(s, COCH_3), 1.24(t, H- β);

^{13}C -NMR (270 MHz, D_2O): δ 17.1 (C- β), 24.9(NHCOCH_3), 57.9 (C-2), 62.9 (C-6), 63.8 (C-6'), 69.1 (C-4'), 71.4 (C- α), 73.8 (C-2'), 74.9 (C-3'), 75.3(C-3), 77.6 (C-5'), 78.2 (C-5), 81.3 (C-4), 103.4(C-1), 105.7 (C-1'), 177.6(NHCOCH_3).

From the above, the product was confirmed to be ethyl N-

acetyllactosaminide represented by structural formula (V):



8-2 Characterization of mannosyl N-acetyllactosaminide (4)

The reaction mixture that seemed to contain mannosyl N-acetyllactosaminide (4) was applied to activated carbon column (ϕ 2.5 \times 25 cm) using activated carbon for chromatography (manufactured by Wako Pure Chemical Industries, Ltd.) and eluted at a flow rate of 1.5 ml/min with a linear gradient of a water/ethanol solution (0 \rightarrow 30%; 2L). The eluate was fractionated into 15 ml fractions and the reaction product was detected by color-development (485 nm) by the phenol-sulfuric acid method. Fractions Nos. 80 to 102 were collected and lyophilized to give about 5 mg of the reaction product. The purity of this product was confirmed to be 99% or more by TLC and liquid chromatography.

A portion of the obtained dried product was analyzed by FAB/MAS and the other portion of the obtained dried product was analyzed by $^1\text{H-NMR}$ and $^{13}\text{C-NMR}$. The results are shown below.

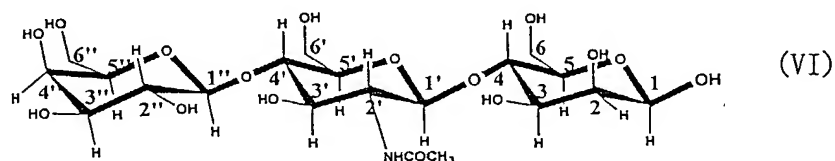
FAB/MAS: (m/z) = 546 ($\text{C}_{24}\text{H}_{46}\text{NO}_{11}+\text{H}$) $^+$, (m/z) = 568 ($\text{C}_{24}\text{H}_{46}\text{NO}_{11}+\text{Na}$) $^+$;

$^1\text{H-NMR}$ (270 MHz, CD_3OD): δ 5.16(d, J 1.6Hz, H-1 α), 4.89(s, H-1 β), 4.58(d, J 8.3Hz, H-1'), 4.47(d, J 7.6Hz, H-1''), (dd, H-2);

$^{13}\text{C-NMR}$ (270 MHz, D_2O): 24.9 (NHCOCH_3), 58.0 (C-2'), 63.2 (C-6'), 62.8 (C-6 α), 62.8 (C-6 β), 63.8 (C-6''), 71.4 (C-4'), 71.8 (C-2 α), 73.0 (C-3 α), 73.5 (C-5 α), 73.5 (C-2 β), 73.8 (C-2''), 74.6 (C-3 β), 74.9 (C-3''), 75.3 (C-3'), 77.5 (C-5 β), 77.6 (C-5''), 78.2 (C-5'), 79.8 (C-4 β), 80.3

(C-4 α), 81.1 (C-4'), 96.4 (C-1 β), 96.5 (C-1 α), 104.2 (C-1'), 105.7 (C-1''), 177.6(NHCOCH₃).

From the above, the product was confirmed to be mannosyl N-acetyllactosaminide represented by the structural formula (VI):



The results of Examples 7 and 8 indicate that lactose and N-acetyllactosaminide themselves are condensed with a compound having an alcoholic hydroxyl group in the presence of the enzyme that cleaves the β -1,4 glucoside bond, so that a corresponding glycoside is produced.

Example 9

First, 1 g of N-acetyllactosaminide (LacNAc), 4.2 ml of glycerol, and 1.4 ml of ethyl acetate were added to 5.6 ml of 100 mg/ml of an enzyme solution, the pH was adjusted to 7.0 with calcium acetate, and then a reaction was performed at 40°C for 48 hours.

The reaction mixture was treated with aniline acetate, and LacNAc was removed. Thereafter, this reaction mixture was loaded to an activated carbon column (ϕ 5 \times 50 cm) using the same support as above to elute an adsorbate at a flow rate of 5 ml/min. A water/methanol solution (linear gradient, 0 \rightarrow 20%; 6L) was used as the developing solvent. The eluate was fractionated into 50 ml fractions and the reaction product was detected by color-development (485 nm) by the phenol-sulfuric acid method. Fractions Nos. 115 to 125 were collected and lyophilized to give about 10 mg of a dried product as the reaction product. The purity of this product was

confirmed to be 99% or more by TLC and liquid chromatography.

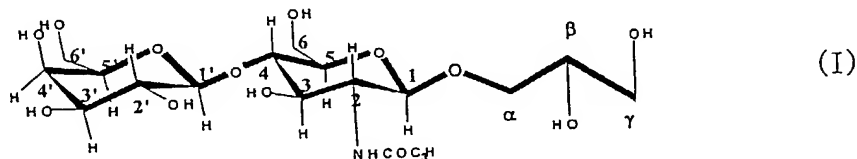
A portion of the obtained dried product was analyzed by FAB/MAS and the other portion of the obtained dried product was analyzed by ^1H -NMR and ^{13}C -NMR. The results are shown below.

FAB/MAS: (m/z) = 458 ($\text{C}_{17}\text{H}_{31}\text{NO}_{13} + \text{H}$)⁺, (m/z) = 607 ($\text{C}_{17}\text{H}_{31}\text{NO}_{13} + \text{Triethanolamine}$)⁺;

^1H -NMR (270 MHz, D_2O): δ 4.55 (d, J 7.3Hz, H-1), 4.47 (d, J 6.8Hz, H-1'), 2.04(s, COCH_3);

^{13}C -NMR (270 MHz, D_2O): δ 25.0 (NHCOCH_3), 57.9 (C-2), 62.9 (C-6), 63.9 (C-6'), 65.2 (C- γ), 71.4 (C-4'), 73.1 (C- β), 73.5 (C- α), 73.8 (C-2'), 75.2 (C-3'), 75.4 (C-3), 77.6 (C-5'), 78.2 (C-5), 81.3 (C-4), 104.2 (C-1), 105.7 (C-1'), 177.6(NHCOCH_3).

From the above, the product was found to be O - β - D- Galactopyranosyl-(1 \rightarrow 4)-O- β -D-2-Acetyl amino-2-deoxyl-Glucopyranosyl-(1 \rightarrow 1)-Glycerol represented by the following structural formula (I).



The obtained compound is a novel substance different from the conventionally reported compounds in the binding pattern between galactose and N-acetylglucosamine, and the binding pattern between N-acetylglucosamine and glycerol.

In the present invention, a novel glycoside in which N-acetylglucosamine is added to glycerol can be obtained. Therefore, known properties and physiological functions and totally novel functions of the conventional glycoside providing glycerol as an aglycon can be expected, and the glycoside of the present invention can be used as a new backbone (raw) material of useful substances such as

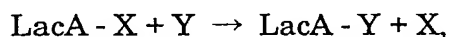
glyceroglycolipids.

Industrial Applicability

The present invention allows lactosyl glycosides and N-acetyllactosaminyl glycosides that are useful as foods, functional food materials, medicines and reagents to be produced under mild conditions, with high yield and in mass quantity and be supplied inexpensively. Furthermore, the lactosyl glycosides and the N-acetyllactosaminyl glycosides according to the present invention contribute to research in medical science and biochemistry.

CLAIMS

1. A method for producing a lactosyl glycoside or an N-acetyllactosaminyl glycoside comprising using a transfer reaction represented by the following formula in a presence of an enzyme having an activity of cleaving a β 1,4 glucosyl bond:



wherein LacA represents lactose or N-acetyllactosaminide; X represents hydrogen (H), a saccharide, a glycoconjugate, or a phenolic compound; and Y represents a compound having an alcoholic hydroxyl group or a phenolic hydroxyl group.

2. The method of claim 1, wherein Y is a compound having an alcoholic hydroxyl group.

3. The method of claim 2, wherein the compound having an alcoholic hydroxyl group is an aliphatic alcohol or a saccharide.

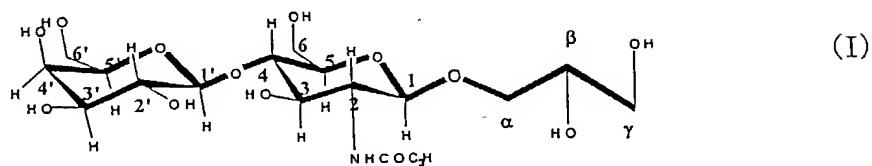
4. The method of claim 2, wherein the compound having an alcoholic hydroxyl group is an amino acid, a peptide or a protein having a serine or threonine residue.

5. The method of any one of claims 1 to 4, wherein X is hydrogen.

6. The method of any one of claims 1 to 5, wherein the enzyme having an activity of cleaving a β -1,4 glucosyl bond is exo-cellobiohydrolase, β -D-glucosidase, and/or cellulase.

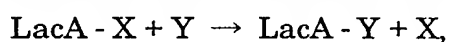
7. Glyceroyl N-acetyllactosaminide (O- β -D-Galactopyranosyl- (1 \rightarrow 4)

-O- β -D-2-Acetylamino-2-deoxyl-Glucopyranosyl- (1 \rightarrow 1) -Glycerol)
represented by the structural formula (I):



ABSTRACT

The present invention provides a lactosyl glycoside or an N-acetyllactosaminyl glycoside useful as foods, functional food materials, medicines, and reagents with high yield and inexpensively by using a transfer reaction represented by the following formula in the presence of an enzyme having an activity of cleaving a β 1,4 glucosyl bond:



wherein LacA represents lactose or N-acetyllactosaminide; X represents hydrogen (H), a saccharide, a glycoconjugate, or a phenolic compound; and Y represents a compound having an alcoholic hydroxyl group or a phenolic hydroxyl group.

Fig. 1

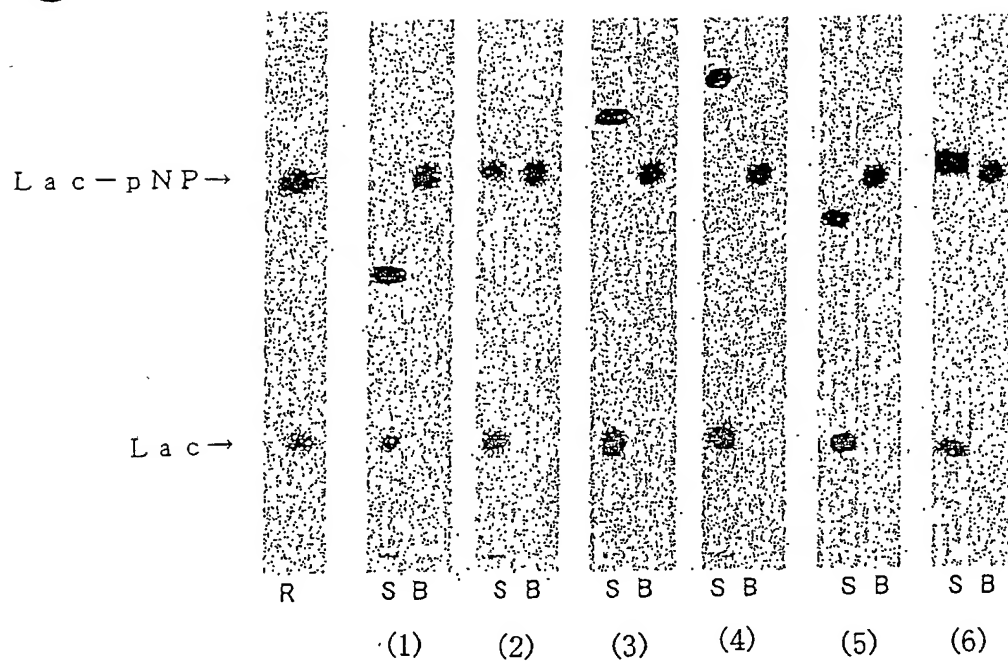


Fig. 2

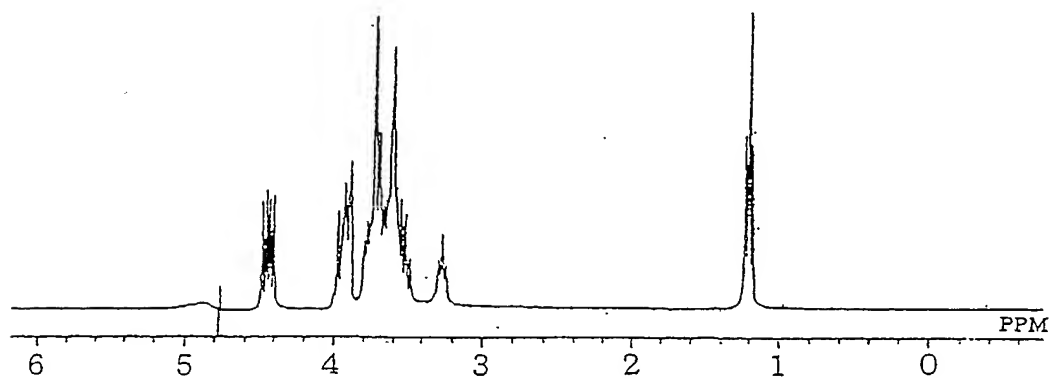


Fig. 3

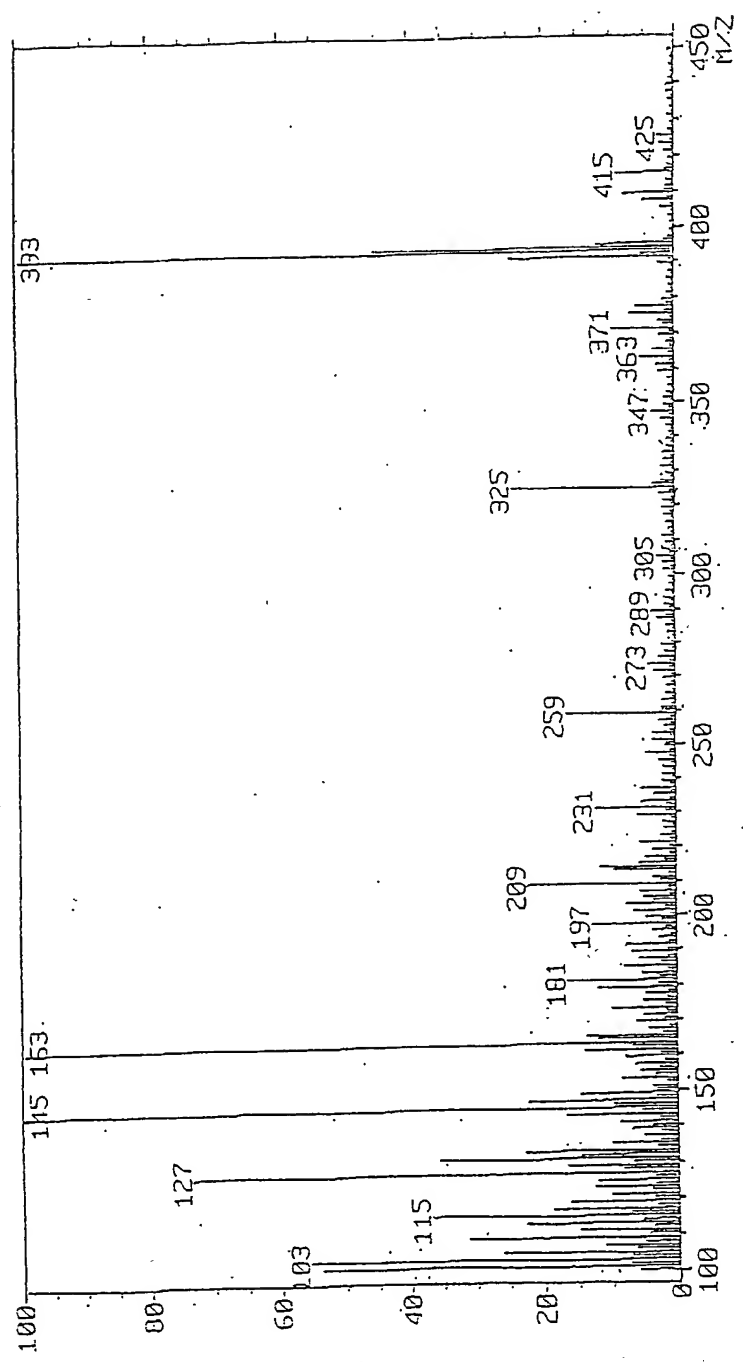


Fig. 4

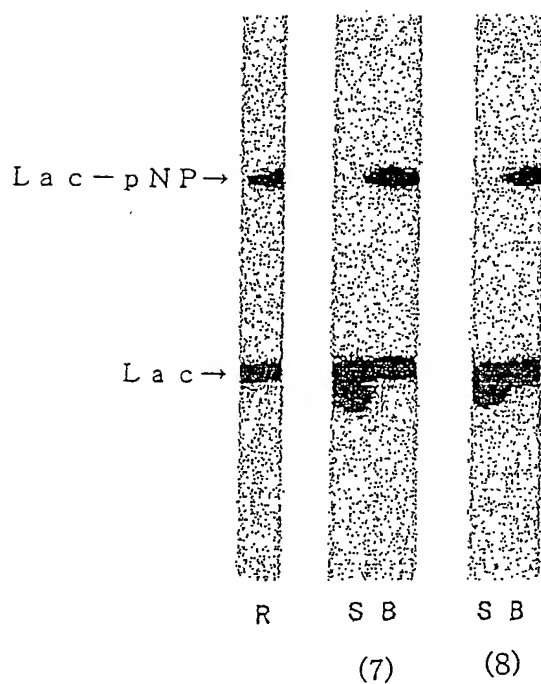


Fig. 5

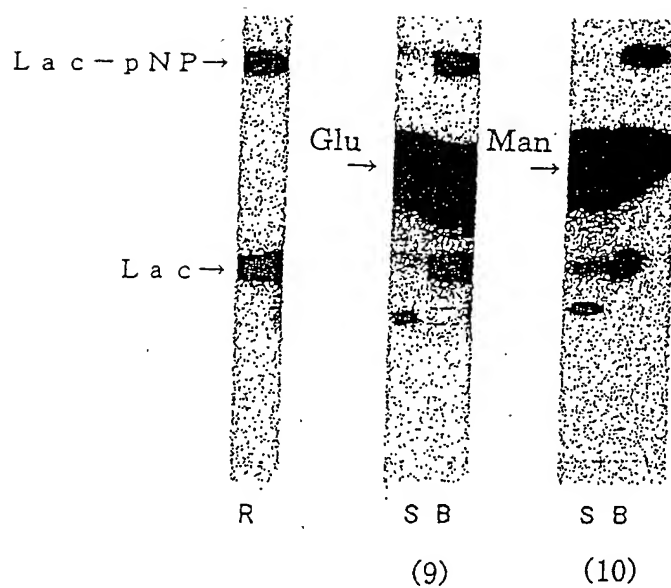


Fig. 6

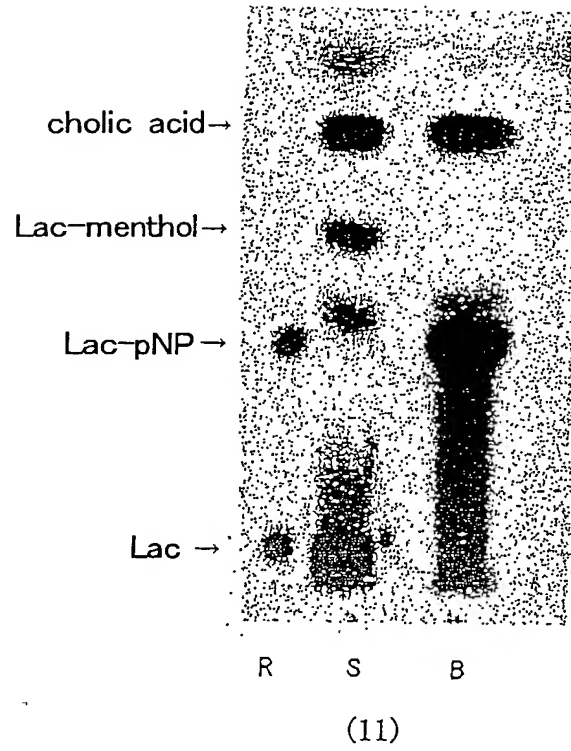


Fig. 7

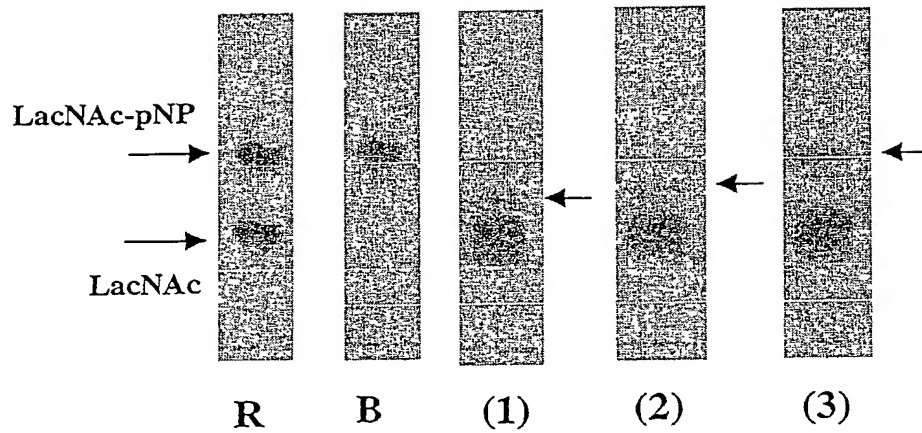


Fig. 8

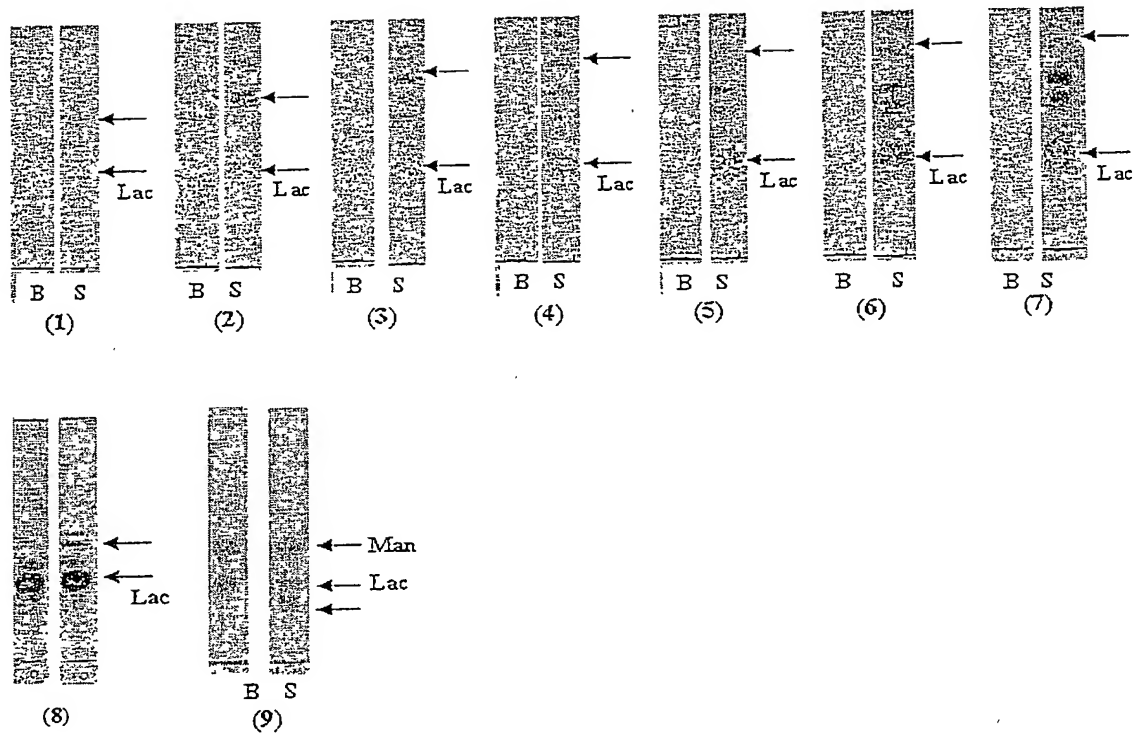
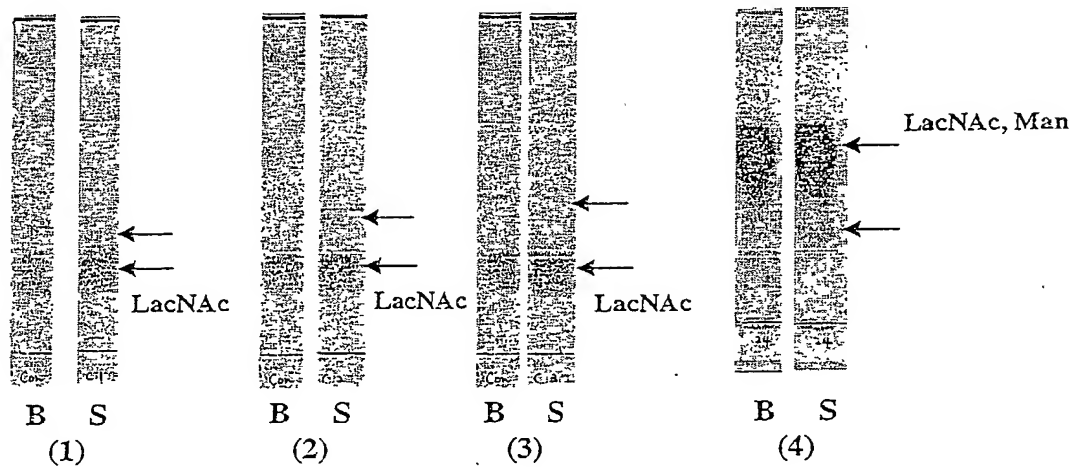


Fig. 9



Declaration and Power of Attorney For Patent Application

English Language Declaration

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name,

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled

Method For Producing Glycosides

the specification of which

(check one)

☐ is attached hereto.

☒ was received on 19 February 2002 as

Application Serial No. 10/049,964

and was amended on 19 February 2002

(if applicable)

☒ was filed as PCT international application

No. PCT/JP00/05576

on 18 August 2000

and was amended under PCT Article 19 on

(if applicable)

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the patentability of this application in accordance with Title 37, Code of Federal Regulations, §1.56(a).

I hereby claim foreign priority benefits under Title 35, United States Code, §119 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

Prior Foreign Application(s)

Priority Claimed

<u>11-233262</u>	<u>Japan</u>	<u>19 August 1999</u>	<input checked="" type="checkbox"/>	<input type="checkbox"/>
(Number)	(Country)	(Day/Month/Year Filed)	Yes	No
<u></u>	<u></u>	<u></u>	<input type="checkbox"/>	<input type="checkbox"/>
(Number)	(Country)	(Day/Month/Year Filed)	Yes	No
<u></u>	<u></u>	<u></u>	<input type="checkbox"/>	<input type="checkbox"/>
(Number)	(Country)	(Day/Month/Year Filed)	Yes	No

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, §1.56(a) which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

(Application Serial No.)

(Filing Date)

(Status)
(patented, pending, abandoned)

(Application Serial No.)

(Filing Date)

(Status)
(patented, pending, abandoned)

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith. (list name and registration number)

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